

# THP1 cells (ATCC TIB-202)

Changes in version 3 (compared to version 2) are in red

Used for: phagocytosis assays

Origin: Human peripheral blood (1 year old boy with acute monocytic leukemia)

Monocytes; suspension cells

Storage: liquid nitrogen vapor phase

| Base medium  | Final conc.          |        |       |
|--|----------------------|--------|-------|
|  |                      |        |       |
| RPMI-1640 <u>ATCC modified</u> (Gibco <b>A1049101</b> ) (stored in fridge) |                      | 500 ml | 50 ml |
| Contains:  |                      |        |       |
| L-glutamine (2 mM)   |                      |        |       |
| Sodium Pyruvate (1 mM)   |                      |        |       |
| Glucose (4.5 g/l)  |                      |        |       |
|  |                      |        |       |
| To make THP1 medium <b>add</b> :   |                      |        |       |
| FBS (heat inactivated, 30 min @ 56°C)*                                     | 5%                   | 25 ml  | 2.5   |
| Pen & Strep 100× (aliquots in -20°C)                                       | 100 u/ml & 100 µg/ml | 5 ml   | 0.5   |
| β-mercaptoethanol, stock 55 mM (sterile, glass bottle, stored at 4°C)      | 0.05 mM              | 500 µl | 50 µl |
|  |                      |        |       |
| <b>Total volume (ml)</b>   |                      | 555.5  | 55.5  |

\* Make sure the serum is warmed up to 37°C before you start inactivating

## Passaging:

Replace medium 3 days per week (afternoons)

Seed: 80,000 – 100,000<sup>#</sup> cells/ml on Friday and 150,000 – 200,000<sup>#</sup> cells/ml on Monday/Wednesday. <sup>#</sup> Range, because it depends on how well they grow.

Cells should **NEVER** exceed 500,000 cells/ml. If this happens, terminate culture.

## PASSAGING

1. **Look at the cells under the microscope.** Check for contamination (fungus, bacteria). Observe morphology and if cells are singular or in clumps. Always use gloves whenever you work with cells.
2. **Count the cells.** Pipette the cells up and down 5–10 × so you will count singular, separate cells only, not clumps. Using the same pipette, transfer a little bit of cell suspension into a new 0.2 ml tube. Use a p20 pipette to transfer 15 µl from this to be used for the cell count. Record number of viable cells/ml, viability % and size of viable cells.
3. **Passage cells.** Take a fraction from your cell suspension (briefly mix by pipetting prior to taking it out) and transfer to a 15 ml tube. This fraction should contain all the cells (total number, not concentration) that you want to seed into the new flask.
  - a. **Mon/Wed.** Seed 150,000 – 200,000 cells/ml (cells will grow for 2 days (48h)).
  - b. **Fri.** Seed 80,000 – 100,000<sup>#</sup> cells/ml (cells will grow for 3 days (72h)).

<sup>#</sup> Range, because it depends on how well they grow.
4. **Centrifuge.** Spin down the cells (use balance tube!) for 5 minutes at 130 × g (acc.9, dec. 5, RT), pipette off the supernatant (take 2 ml from this for mycoplasma testing if applicable) and resuspend the pellet in the desired amount of medium (volume that matches with the size of your culture flask).
5. **Grow** the cells at 37°C and 5% CO<sub>2</sub>. Flasks should be in horizontal position.